ATR-FTIR Spectroscopy Coupled with Chemical and Chemometric Analysis to Distinguish Between Some Sweet Wines

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FTIR Spectroscopy correlated with some chemical characteristics and chemometric analysis have been applied to distinguish between sweet wines obtained from different Romanian varieties and Canadian icewine. Chemical analyses differentiate the two categories of origin in terms of sugar content, acidity and total polyphenol content but are expensive and time-consuming. Principal Component Analysis were performed using different regions of FT-MIR spectra for all wines. Principal Component Analysis of their chemical parameters indicated that the wines can be discriminated based on their different phenolic, carbohydrates, polyols content and geographical origin. FTIR spectroscopy coupled with chemometry is a profitable technique for distinguishing between different wines and validates the results obtained by chemical analysis.

Keywords: sweet wine; mid-infrared spectroscopy; principal component analysis, sugar content, icewine

Grape wine has a very complex composition (hydroxy compounds, carboxylic acids, nitrogenous compounds, aldehydes, vitamins, enzymes and many mineral substances) and a relatively low content of ethyl alcohol (less than 15% V/ V) so it is the healthiest alcoholic drink for the human body. The very complex chemical composition makes it impossible to classify wines according to rigorous scientific criteria. On the other hand, the chemical composition of a wine is unique not only due to its variety footprint, but also to the geoclimatic conditions and winemaking technology. [1, 2].

Sweetness of a wine is determined by the level of residual sugar, remaining after fermentation stops, in balance with alcohol level, acidity and polyphenol content. According to EU regulation 753/2002 a wine containing more than 45g/L residual sugar is considered sweet [3, 4].

The technology of sweet wine production typically involves the dehydration of grape berries in order to increase the sugar concentration. This can be done in different ways. For example, in Tuscany, Italy, the grapes are dehydrated in specially places under shade and controlled temperatures, while in other Mediterranean countries (Cyprus, Greece, Spain, Turkey) are left to air dry, in open spaces. In France (Bordeaux) or Hungary (Tokay) the grapes are attacked by Botrytis cinereal [5].

In cold regions such as those in Canada, Germany, Austria or the United States, the high level of sugar in the famous icewines is getting by freezing the grapes or the grape juice. For these wines, the highly concentrated must is extracted from the frozen berries under pressure condition. The entire harvesting and pressing process must be carried out below -8°C. Juice will have anywhere from 32-46 Brix [6]. Icewine juice is concentrated in all soluble solids, including glucose and fructose, as well as acids and nitrogenous compounds. The management of fermentation biotechnology is very difficult and the fermentation lasts 2-6 months. However, these wines are included in top international level quality premium wines due to their sensory quality and longevity [7].

In Romania, sweet wines with a registered designation of origin (DOC) are obtained by delayed harvest, being classified as: DOC-CMD (full maturity harvest), DOC-CIB (berries ennoblement harvest) and DOC-CT (delayed harvest). Considering the amount of residual sugar, they can be: semi-sweet, with 12.1 - 50 g /L of residual sugar; sweet, with more than 50 g/L of residual sugar. These sweet wines are obtained in different vineyards, from some varieties such as Busuioaca, Muscat Ottonel, Tamaioasa Romaneasca, Grasa de Cotnari, sometimes Chardonnay and Pinot Gris, resulting wines for which sugar and acidity harmonize in the happiest way [8].

For the classification and characterization of sweet wines, numerous techniques and analytical methods can be used: the metallic content [9], volatile fraction [10, 11], aromatic profile [12].

Multivariate analysis has been suggested to differentiate or authenticate wines according to their geographical origin or grape variety using based on the compositional profiles [13].

Some fast, low cost and environmentally friendly methods to characterize the different categories of wine was developed by using FTIR-ATR spectroscopy combined with multivariate analysis [14] or mid-infrared (MIR), near-infrared (NIR), visible (VIS) and ultraviolet (UV) spectroscopy and chemometrics [15, 16].

This study aims to investigate some of the chemical characteristics and to distinguish between sweet wines obtained from different native and Canadian varieties by corelating results of chemical analyses (sugars, *p*H, titratable acidity, total polyphenols and antiradical activity) with FTIR and chemometric analysis.

Experimental part

Wine samples

Eight samples of sweet commercial wine were analysed, five of which were certified in Romania (Cotnari and Mehedinti vineyard) and the other three were icewine produced in Canada.

Table 1 presents some characteristics of wine samples as: name (variety), origin, type of certification and way of obtaining (full maturity harvest-CMD, berries ennoblement harvest-CIB or delayed harvest-CT), year of production, organoleptic properties and ethanol content.

Sugar content

The sugar was determined by the refractometric method using a DigitaleHandrefraktometer DR201-95 which

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Sample	Name (Variety)	Origin (Vineyard) and Certification	Year	Organoleptic properties	Alcohol (% V/V)
1	Grasa de Cotnari (Cotnari 1)	Cotnari DOC-CIB	2008	white, sweet	11.5
2	Tamaioasa Romaneasca (Tamaioasa 1)	Vinju Mare Domains DOC-CMD	2016	white, sweet	13.5
3	Cotnari Inedit- Busuioaca de Bohotin (Busuioaca)	Cotnari DOC-CMD	2017	rose, medium sweet	11.5
4	Grasa de Cotnari (Cotnari 2)	Cotnari DOC-CMD	-	old, white, medium sweet	11.5
5	Tamaioasa Romaneasca (Tamaioasa 2)	Cotnari DOC-CT	2014	white, sweet	11.5
6	Vidal Blanc Aged in oak	VQA Niagara Peninsula –	2016	White, very sweet, very intense and rich flavour	11.0
7	Cabernet Franc	Andrew Peller Signature Series	2016	Red, very sweet, very intense and rich flavour	11.5
8	Vidal		2016	White, very sweet, very intense and rich flavour	11.0

Table 1THE SAMPLES OF WINE ANALYSED

calculate the content in Brix degrees (% sucrose, m/m) based on the index of refraction.

Acidity

The acids play a considerable role in wine quality. There are different organic acids in wines, and each of them contributes differently to total wine acidity or indirectly, to wine colour and stability [17].

Titrable acidity (TA) was determined by titration using a 0.1N NaOH solution (known correction factor), using phenolphthalein as a colour indicator. The results were converted and expressed in g / L tartaric acid using the relationship:

$$TA_{(tartric\,acid,g/l)} = 0,075 \times \frac{V_{NaOH} \times f_{NaOH} \times C_{NaOH}}{V_{sample}} \times 1000$$

where: 0.075 is the amount of tartaric acid corresponding to 1 meq H⁺/L, V_{NaOH} is the volume of NaOH solution with the concentration C_{NaOH} and correction factor f_{NaOH} used in the titration and V_{sample} is the volume of sample taken in the analysis.

Actual acidity or real acidity expressed by *p*H was measured using an InoLab 730 pH-meter

Total polyphenols content

The total polyphenols content (TPC) was determined by the Folin-Ciocalteu method, that involved the reduction of the F-C reagent by the phenolic compounds in wine with the formation of a blue complex which has a maximum of absorption at 760 nm. The absorbance was interpolated and expressed in gallic acid on a calibration curve obtained with a series of 12.5 - 150 μ g/mL galic acid solution and describe by a linear regression equation. The results were obtained as an average of three consecutive determinations and read at a UV-Vis Jasco 730 spectrometer.

By multiplying the absorption values by 20, in the case of white wines, and by 20 x dilution factor, for the red ones, we have the total polyphenols index (TPI) that allows to classify the wines in: supple taste wines (TPI < 30), well-formed wines (30 < TPI < 50) and astringent, with excess of polyphenols wines (TPI > 50) [8].

Antiradical power

The evaluation of the antiradical power of the wine was made in comparison with 2,2-Diphenyl-picrylhydrazyl (DPPH), one of the most stable free radical organic compounds. It accepts H from the antioxidants present in the sample and the antioxidant effect is proportional to the disappearance of the DPPH radical. DPPH has a strong peak absorption at 517 nm (violet). The colour changes to yellow due to DPPH H formation. The antioxidant effect can be evaluated by decreasing absorbance at 517 nm [18].

The DPPH (394.32 gram /mol) methanolic solution with a concentration of $6x10^{-5}$ mol/L (2366x10⁻⁵g/L) in methanol was obtained and then, the reference solution was prepared by diluting 3900 µL with 100 mL methanol. Radical reaction was initiated by transferring to 25 mL of each wine sample 3975 µL of reference solution (dilution ratio 1: 160). For each sample, the absorbance value was measured after 15 min of incubation in the dark and at the room temperature, at 517nm, on a UV-Vis Jasco 730 spectrometer.

Percent inhibition of DPPH. (I%) was calculated from the decrease in absorbance with the relation:

$$I\% = \frac{A_{blank} - A_{sample}}{A_{blank}} \times 100$$

where: A_{blank} is the absorbance for the blank (reference solution of ethanol- DPPH ethanolic solution) and A_{sample} is the absorbance for the sample mixed with DPPH reference solution.

The ATR-FTIR spectral analysis

The ATR-FTIR spectra were recorder in a range between 4000-400 cm⁻¹ using a FTIR Jasco 6300 spectrometer, detector TGS, apodization Cosine. An ATR accessory equipped with a diamond crystal (Pike Technologies) was used for sampling. Each sample $(10\mu L)$ was spread uniformly through on the surface of the diamond ATR crystal using a micropipette.

At the same time, the viscous residues were also obtained after 1 mL of each sample was dropped on a pan and volatilized for 8 h. For each viscous and liquid wine samples the IR spectra were recorded and analysed for classification.

The spectral data were processed with JASCO Spectra Manager II software. Liquid samples of wine, without any preparation, were scanned at 4 cm⁻¹ resolution, accumulation: 100 scans. For each subsample, three replicate spectra were recorded to ensure the spectral reproducibility and assess analytical precision and the average spectrum was done. Background reference spectra were recording using air after every sample to minimize the interference due to carbon dioxide and water vapor in the atmosphere. Between measurements, the ATR crystal was carefully cleaned using acetone then dried with a soft tissue.

Data Analysis

Infrared Spectra were exported from Spectra Manager, in ASCII (dx) format, into the Unscrambler Software (Edition X 10.3, Camo. Oslo Norway) for chemometric analysis. Spectra were pre-processed using the secondderivative transformation, the Savitzky-Golay derivation. The use of spectra derivatives with Savitzky-Golay algorithm as a chemometric pre-processing technique is widely reported in most classification based on FTIR spectroscopy [12-14, 19-21]. The principal component analysis (PCA) model was developed using cross validation. PCA was performed both on the entire spectral range (4000 to 400 cm⁻¹), and on the MIR 'fingerprint' (i.e. 1800 to 700 cm⁻¹), on the peak areas corresponding to regions 940-600 cm⁻¹ (1), 1000-1120 cm⁻¹ (2) and 3000 -2800 cm⁻¹ (3).

Results and discusions

Table 2 comprise the values obtained for the chemical parameters investigated for the eight wine samples.

Analysing the sugar content, Icewine samples are found to have the highest values (27-29 % Brix), practically double quantities than the ones contained in local wines (10-14 % Brix). These values are positively correlate with the titratable acidity (TA) expressed in tartric acid equivalents. For this parameter higher values also characterize the Canadian wine. A minimum value (4,53 g tartric acid eq /l) is obtained for sample 2, Tamaioasa Romaneasca from Vinju Mare Domains which is the wine with the highest alcohol content (13.5%).

There is no proportionality between titratable acidity and the *p*H values indicating the degree of ionization of acids and acid salts of wine. Concentration of free hydrogen ions is higher for all the autochthonous varieties (sample 1-5).

Polyphenols content are also important among the chemical compounds present in wine not only because of their influence on colour and taste but also due to their antioxidant activity and benefices on human health [22, 23]. These molecules confirmed to possess a freeradical scavenging activity, so that the total polyphenols content can be correlated with the antiradical power evaluated by the DPPH assays. Recent studies have shown that polyphenols are active in inhibiting radicals, while sugars are effective in inhibiting assays mediated by hydroxyl radical formation. Sugars display antioxidant capacities that cannot be underestimated, and their action can involve polyphenols especially in samples with more than 500 g/L sugar concentration [24].

In the analysed samples, the Cabernet Franc icewine variety stands out with a much higher content of

polyphenols and the highest DPP inhibition power. This variety, a well-formed wine (TPI > 30), with a brilliant ruby hue, is considered very rare and precious and is specific to the Ontario-Canada area. For the other samples, the values are comparable and place the wines in the category of supple to taste (TPI < 30).

FT-MIR spectral fingerprinting of wine samples

The ATR-MIR spectra of commercial wines showed moderate to strong absorbance peaks at 1044, 1085, 1636-1638 and around at 3270 cm⁻¹ (fig. 1) with peaks at 3270 and 1636-1638 cm⁻¹ corresponding to the O-H stretching and bending respectively, associated with water [25, 26].

In all samples, similar spectral features were generally obtained. Figure 1 exhibits the general FT-MIR spectra of all wine residue samples.

In order, to get an overview of the compositional differences that occurred in these eight samples of wine an overlapping image of the of Romanian and Canadian wine samples was taken and it is presented in figure 2. Various spectral bands occur in the region from 1800-700 cm⁻¹, which is part of the fingerprint region, such as those corresponding to the vibration of the C-O, C-C, C-H and C-N bonds. It was possible to consider a few specific regions which can be useful for wines' characterization [27-29]. Region 1 (690-940) is responsible for phenols, region 2 (1000-1120 cm⁻¹) is responsible to carbohydrates (glucose, fructose and oligosaccharides), and region 3 (2800-3000 cm⁻¹) for polyols (mainly glycerol). Table 3 includes the FT-MIR absorption wave numbers,

specific to the 3 regions of each wine fingerprinting.

All spectra were characterized by a similar profile. No visual differences were observed between the MIR spectra of wine samples analysed. It was observed that water and ethanol absorption peaks dominate the spectrum. Chemometric analysis allow the differentiation of wines.

The chemometric analysis. Principal Component Analysis (PCA)

The Principal Component Analysis (PCA) is a wellestablished analysis technique which works by finding the correlation between a set of variables and then creating a new set of uncorrelated variables named principal components (PCs). PCA was performed on the MIR spectra to examine qualitative differences within the set of sweet wines related to the two production systems [14, 29, 30].

Figures 3-6 present the results of PCA analysis and scores. PCA was carried out between the domain of the selected spectral region 1800-700 cm⁻¹ for wine (discriminate sweet wine types: Romanian and Canadianfig. 3). The first three principal components (PCs) represent 97% of the total variance (PC1 = 72\%, PC2=22\%; = 3\%) and PC3 = 3%). This indicates that these three components were sufficient to provide a clear separation

Sample	Sugar	Acidity		Total polyphenols		1%]
	% BRIX	pH	Titrable acidity (TA)	Total polyphenols content	TPI	DPPH	
			g tartric acid /l	(TPA)		inhibition	
				µg galic acid /ml			Table 2
1	13.6	2.99	4.84	46.0455± 0.0283	13.53	68.42	THE SUGAR, ACIDITY AND
2	12.0	2.88	4.53	33.1692 ± 0.0405	10.08	52.58	TOTAL POLYPHENOLS
3	10.6	2.83	5.36	51.2353 ± 0.0533	14.92	76.94	CONTENT IN WINE SAMPLES
4	11.1	2.86	5.51	40.0088 ± 0.0695	11.91	56.44	
5	13.8	2.76	5.74	55.2947± 0.0508	16.01	60.47	1
6	29.0	3.44	7.12	61.5184 ± 0.0363	17.68	54.45	1
7	28.1	3.62	7.35	125.0095 ±0.0510	33.68	82.51	1
8	27.7	3.59	6.90	57.2224 ± 0.0156	16.53	51.83	1

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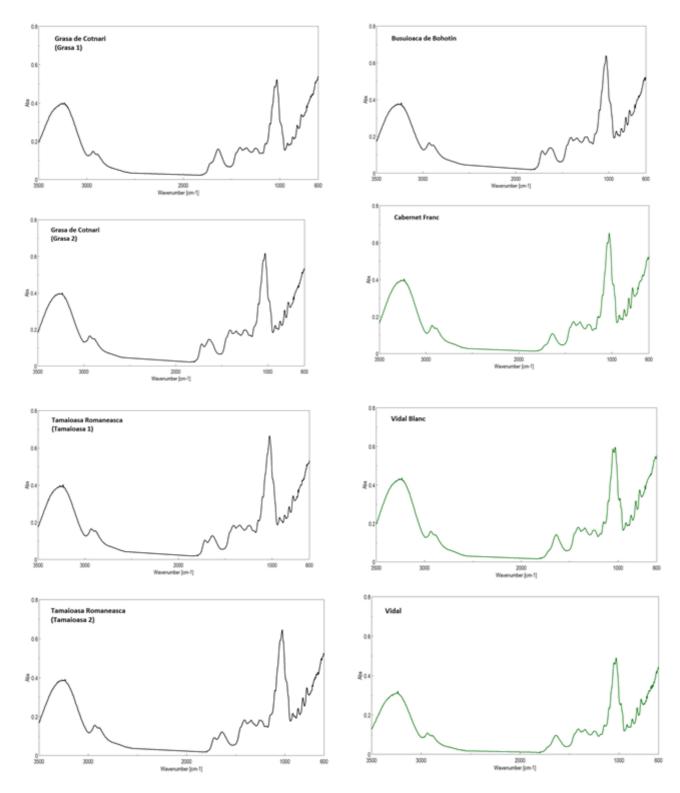


Fig. 1. ATR-MIR spectra registered for all 8 sweet wine samples, in the region 600-3500 cm⁻¹

between the groups. The Romanian wine varieties are located on the right of the plot, while Canadian sweet wines are located on the left of the graph.

Figure 4 represents the scores for the region 600-940 cm⁻¹, specific to for phenolic compounds (chemical compounds that affect the taste, color and mouthfeel of wine). In this case, the first principal component (PC1) explained 74% of the variability, the second principal component (PC2) explained 23% and the third principal component (PC3) with 2% of the variability, together explaining 99% of the whole variability of wine samples and suggesting a good clustering based on acidity. The type of wine separates: in the right are the Romanian wines (Grasa de Cotnari - Grasa 1, Tamaioasa Romaneasca Vanju

Mare - Tamaioasa 1, Busuioaca de Bohotin - Busuioaca, Grasa de Cotnari - Grasa 2 and Tamaioasa Romaneasca Cotnari- Tamaioasa - 2) and in the left part of diagram are Canadian wines (Vidal Blanc Aged in Oak, Cabernet Frans and Vidal). Moreover Grasa de Cotnari 1 and 2 are in the lower part of the right of plot and Tamaioasa 1 and 2 and Busuioaca are in the positive quadrates of both PC1 and PC2. Cabernet Franc wine is separated from the other canadian wines. These are in concordance with total polyphenols values obtained by the Folin-Ciocalteu method (table 1).

Figure 5 which represents the scores for the region 1000-1120 cm⁻¹, specific to carbohydrates derivatives, revealed a good clustering of wines based on sweetness index. The

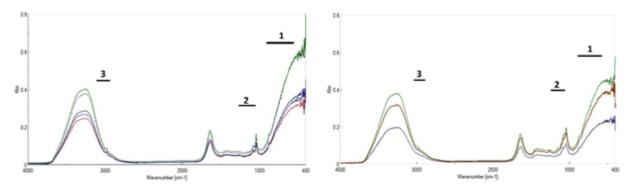


Fig. 2. Overlapped ATR-MIR spectra for the five sweet Romanian commercial wine samples (left) and three Canadian Icewine (right)

Table 3
ATR-MIR WAVE NUMBERS SPECIFIC TO THE REGIONS OF EACH WINE FINGERPRINTING

Sample of wine	1 (600-940 cm ⁻¹)	1 (1000-1200 cm ⁻¹)	3 (2800-3000 cm ⁻¹)
Grasa de Cotnari (Grasa 1)	677, 719, 775, 817, 864,918	1029, 1096	2888, 2936
Tamaioasa Romaneasca (Tamaioasa 1)	675, 689, 719, 732, 774, 816, 864, 917	1025, 1099	2888, 2935
Cotnari Inedit- Busuioaca de Bohotin (Busuioaca)	668, 677, 719, 776, 816, 863, 899. 916	1027, 1099	2887, 2934
Grasa de Cotnari demidulce (Grasa 2)	669, 719, 775, 816, 862, 900, 917	1028, 1100	2886, 2934
Tamaioasa Romaneasca (Tamaioasa 2)	664, 677, 691, 719, 775, 816, 864, 901, 918	1026, 1100	2883, 2934
Vidal Blanc Aged in oak	642, 659, 719, 776, 816, 862, 891, 916	1029, 1051, 1100	2888, 2935
Cabernet Franc	612, 624, 659, 719, 776, 816, 864, 917	1027, 1046, 1100	2889, 2935
Vidal	620, 647, 666, 697, 719, 729, 749, 776, 816, 864, 917	1028, 1050,1101	2884, 2937

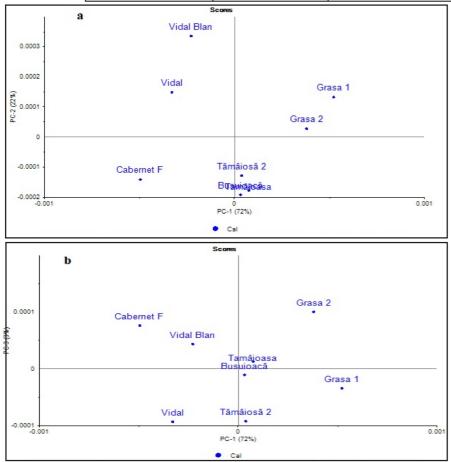
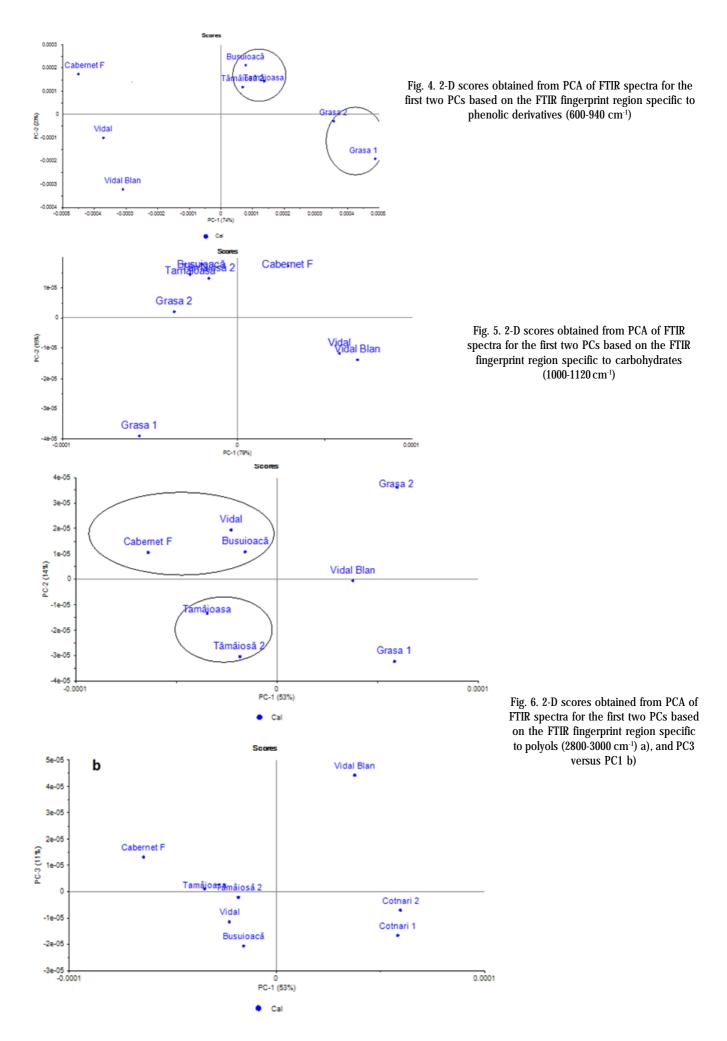


Fig. 3. 2-D scores obtained from PCA of FTIR spectra for the first two PCs based on the FTIR fingerprint region (700-1800 cm⁻¹) a), and PC3 versus PC1 b)



first PC explains 79% while the second and the third principle component 2 and 2% respectively of the total variance. The type of wine separates: in the right of the diagram are the Canadian sweet wines. These are in concordance with sugar content values presented in table 1.

Figure 6 represents the scores for the region 2800-3000 cm⁻¹, specific to polyols (mainly glycerol). The first PC explains 53% while the second and the third principle component 14% and 11% respectively of the total variance. Vidal Blanc Aged in oak is separated from the rest two Canadian wines. This result may be due to processes that occur during wine aging, such as: the release of some compounds from the oak wood into the wine; chemical reactions involving both wine and wood compounds as well as oxygen permeation through the wood; evaporation of volatile compounds with the concentration of non-volatile ones [31, 32].

Polyols especially glycerol afford *fullness to wine* and decrease the perception of acidity, particularly in dry white wines. Also glycerol significantly contributes to the complexity of wine and the length of the finish. Its contribution to sweetness was considered significant, but only when present at high concentrations [33].

Conclusions

This study demonstrated the capacity for ATR-MIR spectroscopy (combined with multivariate analysis) to broadly classify wines. The results demonstrated qualitative compositional differences between sweet Romanian and Canadian wine, that can be observed by MIR spectroscopy and used to distinguish wines, following PCA.

The chemometric instruments are rapid and allow an accurate separation of the wine by the sugar content, phenols, polyols, with very good results from the spectrum of a drop of wine in the absence of the chemical analysis of the sample necessary for these determinations.

The chemical analyzes show that the Romanian sweet wines and Canadian icewines are distinguished by the sugar content, acidity and the total content of polyphenols. The differences are particularly remarkable in terms of sugar content where the values are double in icewines versus the Romanian ones. All wines demonstrate antiradical activity without major differences between the two groups of origin. However, chemical analyzes are laborious, time consuming and expensive. In contrast, ATR-MIR spectroscopy combined with multivariate analysis proved to be a quick and effective technique to distinguish between different samples of sweet wine.

The separation of wines by chemometry according to the content of carbohydrates, phenols, polyols is in good concordance with that obtained by chemical methods

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